

Impact of *lpxM* deficiency on the drug sensitizing activity of the efflux pump inhibitor PA β N

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Background: PA β N (phenylalanyl arginyl β -naphthylamide) is an early agent shown to inhibit RND-type efflux pumps of Gram-negatives such as AcrAB-TolC from *Escherichia coli* [1]. However, an additional permeabilizing activity has been reported [1, 2]. Previously, we had selected an *E. coli* mutant (C5/1/17) showing partial resistance to the drug sensitizing activity of PA β N but without any mutations in *acrAB-tolC* [3]. In this study, we intended to elucidate the mode(s) of action of PA β N.

Methods: An *acrAB-tolC* overexpressing *E. coli* was subjected to error-prone PCR based in-vitro random mutagenesis targeting *acrB*. Mutants were selected in a single step on clarithromycin in the presence of PA β N. *acrB* alterations from mutants with confirmed PA β N resistance were reconstructed. Whole genome sequencing was applied in the case of lacking *acrAB-tolC* mutations associated with PA β N resistance. Phenotypic characterization was done by MIC testing with and without EPIs or membrane permeabilizers, and by dye accumulation and efflux assays.

Results: From targeted (*acrB*) mutagenesis, we obtained a mutant CP1 that, similar to C5/1/17 (obtained from serial selection), showed resistance to PA β N activity. We detected 4 mutations in *acrB* of CP1 (V129I, L270V, T495S, A873V), but their reconstruction within the parent did not reveal any association with PA β N resistance. However, this was maintained when replacing mutated *acrB* by wild-type *acrB* in CP1. By WGS sequencing we discovered loss-of-function mutations in *lpxM* of CP1 as well as of C5/1/17 (stop codon and frameshift, respectively). *lpxM* is coding for a myristoyl acyltransferase and its loss is known to result in pentaacylated lipid A. Interruption of *lpxM* within the parental strain confirmed its impact on PA β N activity. We found largest effects with drugs known to display high synergy with PA β N (8-16 fold decreased PA β N activity with novobiocin, rifamycines, macrolides). The permeability of the mutant for some agents appeared increased (nitrocefin, rifampin, polymyxin B nonapeptide), but only marginal for macrolides, rifaximine, colistin, and PA β N itself (\leq 2-fold, MIC parent / MIC mutant). Real-time efflux assays confirmed unimpaired AcrAB-TolC functionality. Interestingly, Δ *lpxM* was also associated with decreased drug sensitizing activity of colistin, but not of NMP and MBX2319.

Conclusions: We discovered a target of PA β N in the outer membrane proving an alternative mode of action that contributes to the drug sensitizing activity of this EPI.

References

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